THE EFFECT OF IMMUNOSTIMULANTS FROM NATURAL INGREDIENTS ON VANAMEI SHRIMP (*LITOPENAEUS VANNAMEI*) IN INCREASING NON-SPECIFIC IMMUNITY TO FIGHT DISEASE

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ABSTRACT

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Keywords :

Litopenaues vannamei, immunostimulants, nonspecific immunity, natural ingredients, shrimp disease Vanamei shrimp Litopeneus vannamei is a commodity with high economic value. Shrimp is Indonesia's main export commodity. The high value of sales and production of vanamei shrimp is constrained by the many disease attacks in the vanamei shrimp farming industry. An alternative solution that has been continuously researched for several years to combat shrimp disease is using immunostimulants from natural ingredients. Using natural ingredients for immunostimulants can increase the non-specific immunity of vanamei shrimp to ward off disease and be environmentally friendly. Immunostimulants can be administered orally, by immersion, or by injection. Immunostimulants from ingredients such as spirulina platensis, natural polysaccharides from seaweed such as carrageenan, sodium alginate, and fucoidan, extracts from red seaweed, as well as the use of polysaccharides from the bacterium Nodulisporium sp. KT29, Trichoderma sp. able to increase the non-specific immune system in L. vannamei shrimp. These immunostimulants increase non-specific immunity, such as total hemocyte count, phagocytosis activity, phenoloxidase, phagocytic, respiratory burst, superoxide dismutase, and total plasma Immunostimulants from these protein. natural ingredients can also fight infectious disease attacks.

INTRODUCTION

The vannamei shrimp *Litopeneus vannamei* is a commodity with high economic value and is Indonesia's main export commodity. The export value of this commodity reaches USD 1 billion or 40.1% of the total export value (DITJEN PDSPKP, 2021). Vannamei shrimp originated from Hawaii and Florida and were later introduced to Indonesia. They are widely cultivated because they have several advantages, including fast growth and low feed conversion value, and are very suitable for cultivation in ponds with high stocking densities (Lestari et al., 2018).

Even though the export value of vanamei shrimp is high, it still has problems in the form of many disease attacks on the vanamei shrimp farming industry. Disease caused by *Vibrio*

harveyi resulted in the death of around 80-100% vannamei shrimp (Widanarni et al., 2012). Vibriosis is a shrimp disease problem not only in Indonesia but also a global problem (Cao et al., 2015). Another disease that attacks the shrimp farming industry is *infectious myonecrosis* (IMN) which caused losses of up to 300 billion in Indonesia in 2009 (KKP, 2010). IMN disease causes the death of 40-70% of vanamei shrimp (OIE, 2009). In addition to diseases caused by bacteria, there are also diseases caused by viruses, namely the *White Spot Syndrome Virus* (WSSV), *Yellow Head Virus* (YHV), and *Taura Syndrome Virus* (TSV), which are considered the most dangerous pathogens in paneid shrimp because they cause losses. Large ones (Loh et al., 1997; Wang et al., 2007).

Many vannamei shrimp cultivators cope with disease attacks by using antibiotics (Defoirdt et al., 2011). Apart from antibiotics, some use vaccines or chemotherapy to treat diseases in vanamei shrimp but are considered ineffective (OIE, 2009). However, the use of this antibiotic raises a new problem, namely the resistance of the bacteria that attack vanamei shrimp to antibiotics, which threatens the shrimp's health and the health of the shrimp consumers.

The vanamei shrimp *L. vannamei* has a very primitive immune system compared to fish and other vertebrates (Kwang, 1996). The crustacean shrimp group only has innate or nonspecific immune defense abilities and does not produce specific antibodies (Anderson, 1992). This non-specific immune system helps shrimp to fight disease. This non-specific defense can only be stimulated daily, so the stimulation should be done periodically and continuously, either by oral or immersion which is a treatment with little risk.

An alternative solution that has been continuously researched for several years to tackle vanamei shrimp disease is the use of immunostimulants from natural ingredients (Febriani, 2013). According to Sakai (1999), immunostimulants can increase non-specific immunity in shrimp. Improving the non-specific immune system is positively correlated with increasing the health of shrimp to fight disease. Using natural ingredients for immunostimulants can increase shrimp's non-specific immunity to ward off disease and be environmentally friendly (Zahra et al., 2017). Giving immunostimulants from natural ingredients to vanamei shrimp is expected to solve disease problems in vanamei shrimp by increasing non-specific shrimp immunity and being environmentally friendly.

DISCUSSION

The use of natural ingredients as immunostimulants to treat disease in *L. vannamei vanamei shrimp* has been widely studied lately. Using natural ingredients as immunostimulants are considered because they are environmentally friendly and can potentially increase non-specific immunity. Natural ingredients that are widely used come from the seaweed group (Muahiddah & Sulystyaningsih, 2021). Many groups of seaweed are used as immunostimulants from red seaweed, namely gracilaria, brown seaweed, sargassum, and laminarin. Apart from seaweed, there are also natural ingredients from herbal ingredients, namely black cumin and garlic, as immunostimulants in improving health conditions, quality of life of fish, growth and increasing immunity and enabling fish to cope with stress caused by artificial infections (Ahmed et al., 2008).

Seaweed cell walls are rich in polysaccharides which can be helpful as immunostimulants, anticoagulants, antioxidants, antibacterial, antiviral, anti-cancer, and immune-modulating activation (Wijesekara et al., 2011). Red, green, and brown seaweeds are a source of immunostimulants because they contain bioactive that produce secondary metabolites with a broad spectrum of activity and can enhance shrimp non-specific immunity

(Febriani et al., 2013). Polysaccharides from seaweed often used are carrageenan (red seaweed), fucoidan, and sodium alginate (brown seaweed). In addition to polysaccharides from seaweed, polysaccharides from bacterial walls such as *Nodulisporium* sp. KT29 and *Trichoderma* sp.

Immunostimulants can be administered orally, by immersion, or by injection. Research on determining the dose and frequency is still being carried out to determine the most effective way to use immunostimulants. The use of immunostimulants in vannamei shrimp (*L. vannamei*) in the following references:

Immunostimulants	Method	Results	Resistant to disease	Reference
Kappa-Carageenan	Oral	Total Haemocyte Count (THC)	Infectious	Febriani et al.,
		Phagocytosis	myonecrosis	2013
		Phenoloxidase	(IMN)	
		Survival Rate (SR)		
Extract	Oral	Total Haemocyte Count (THC)	White spot	Zahra et al., 2017
Gracilaria verrucosa		Respiratory burst (RB)	syndrome virus	
		Phenoloxidase	(WSSV)	
		Survival Rate (SR)		
Fucoidan	Oral	Total hemocyte count (THC)	White spot	Setiawan, 2019
		Phagocytosis (AF) activity	syndrome virus	
		Phagocytosis index (IF)	(WSSV)	
		Superoxide dismutase (SOD)		
		activity		
		Phenol oxidase (PO) activity		
		Total plasma protein (TPP)		
Sodium Alginate	Oral	Total hemocyte count (THC)	-	Surur, 2018
		Phagocytosis (AF) activity		
		Phagocytosis index (IF)		
Spirulina platensis	Oral	Total hemocyte count (THC)	Vibrio harveyi	Ghowina, 2018
		Differential total hemocytes (DHC)		
		Phagocytosis (AF) activity		
<i>Nodulisporium</i> sp. KT29	Oral	Respiratory burst (RB)	Vibrio harveyi	Wahjuningrum et
		Phenoloxidase		al., 2020
		Survival Rate (SR)		
Trichoderma sp.	Injection	Total hemocyte count (THC)	Vibrio harveyi	Pebrianto, 2009
		Survival Rate (SR)		
		Phagocytic index		
		Phenoloxidase activity		

Polysaccharides from seaweed or bacterial walls can stimulate non-specific immunity, such as phagocytic activity, and respiratory burst through interaction from the receptor surface (Siregar et al., 2012). Polysaccharides increase the total hemocyte count. The total hemocyte count is a non-specific indicator of both humoral and cellular defenses. Change in total hemocytes is an indicator not only of stress but of the health status of the shrimp. In addition, hemocytes are involved in synthesizing and releasing essential molecules such as α -2-macroglobulin (α 2M), agglutinins, and antibacterial peptides as a reaction of the body's defense in shrimp (Rodriguez & Moullac, 2000). Total hemocytes are closely related to proPO stimulation to induce phenoloxidase activity. Phenoloxidase activity indicates the ability of vannamei shrimp to recognize foreign bodies that enter the body (Garcia-Carreno et al., 2008) and the activity of the shrimp's body defense (Costa et al., 2009).

Hemocytes play an essential role in the immune system of shrimp. First, hemocytes can remove foreign particles or disease in the hemocoel through phagocytosis, encapsulation, and nodular aggregation mechanisms (Soderhall & Cerenius, 1992). Both hemocytes in shrimp play a role in wound healing through cellular aggregation and initiation of the coagulation process through the release of factors required for plasma gelation (Johansson & Soderhall, 1989; Omori et al., 1989; Vargas-Albores et al., 1998), and the release of prophenoloxidase proPO system (Johansson and Soderhall, 1989 Hernandez-Lopez et al., 1996).

Melanization is essential to the shrimp's immune defense system against pathogens. Phenoloxidase is an enzyme that plays a role in the process of melanization. This enzyme is produced through the proPO (prophenoloxidase) system, which is activated in the presence of an immunostimulant. ProPO plays an essential role in introducing foreign bodies, including phagocytosis, melanization, production of cytotoxic reactants, particle encapsulation, and formation of nodules and capsules (Amparyup et al., 2013). Activation of the proPO system induces the production of melanin, a brown pigment responsible for various processes, such as activation of foreign particles, protecting their distribution in the host's body, and repairing cuticle damage.

Phagocytosis is the most common response in cell protection. Through phagocytosis, the cellular immune response in shrimp responds to foreign bodies. Advantages in phagocytosis activity in shrimp also occur in several post-challenge test studies as a shrimp defense mechanism (Jasmanidar, 2009; Febriani et al., 2013). During phagocytosis, particles or microorganisms enter the cell and form digestive vacuoles called phagosomes. The mechanism of particle removal by phagocytic cells, called the respiratory burst, involves the release of degradative enzymes into the phagosome (an oxygen-dependent killing mechanism) and the production of ROI (reactive oxygen intermediates) (Rodriguez & Moullac, 2000). An increase in the respiratory burst is correlated with an increase in phagocytosis, viral particles are recognized by receptors on the cell surface and are engulfed by cells that reprepare the cytoskeleton for phagosome formation. In the first stage, phagosomes undergo maturation through cleavage and fusion with lysosomes and become mature α phage-lysosomes. Viruses in phage-lysosomes will be destroyed by low pH conditions, hydrolysis processes, and radicals (Xu et al., 2014).

Wongprasert et al. (2013) stated that sulfated galactan could stimulate the shrimp immune system, possibly mediated by the interaction between sulfated galactan and receptors on the hemocyte surface. The existence of a relationship between galactan sulfate and these receptors will activate the signaling process of increased hemocyte proliferation and stimulation of immune system activity (Zahra et al., 2017).

Li and Xiang (2013) stated that shrimp receptors, namely pattern recognition receptors (PRRs), play an essential role in the shrimp immune system and consist of lipopolysaccharide, -1,3-glucan binding protein (LGBP), and toll receptors. Pathogen recognition by PRRs is triggered by activation via the serine protease pathway and finally cleaved to proPO to produce phenoloxidase. Galactan sulfate in *Gracilaria verrucosa extract* containing 1,3-glucan interacts with LGBP in the hemocyte membrane, which activates the phenoloxidase enzyme. Other receptors that play an essential role in the shrimp immune system are toll-like receptors (TLRs).

CONCLUSION

Immunostimulants from natural ingredients such as *spirulina platensis*, polysaccharides from seaweed such as carrageenan, sodium alginate, and fucoidan, extracts from red seaweed, as well as the use of polysaccharides from the bacterium *Nodulisporium* sp. KT29, *Trichoderma* sp. able to increase the non-specific immune system in *Litopeneus vannamei shrimp*. These immunostimulants increase non-specific immunity, such as total hemocyte count, phagocytosis activity, phenoloxidase, phagocytic, respiratory burst, superoxide dismutase, and total plasma protein. Immunostimulants from these natural ingredients can also fight infectious myonecrosis (IMN), *White spot syndrome virus* (WSSV), and *Vibrio harveyi bacteria* by increasing the survival rate of vannamei shrimp that are not given immunostimulants.

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REFERENCES

- Anderson, DP, 1992. Immunostimulants, adjuvants, and vaccine carriers in fish: application to aquaculture. Annual Review of Fish Diseases 2, 281-307.
- Ahmed SD, Aly SM, George J, Yasser AH, Mohamed FM. The effect of garlic, black seed and Biogen as immunostimulants on the growth and survival of Nile tilapia, *Oreochromis niloticus* (*Teleostei cichildae*) and their response to artificial infection with *Pseudomonas fluorescence* . African Journal of Aquatic Science. 2008;33(1):63-68.
- Amparyup P, Charoensapsri W, Tassanakajon A. 2013. Prophenoloxidase system and its role in shrimp immune responses against major pathogens. Fish and Shellfish Immunology 34: 990–1001.
- Cao, H., An, J., Zheng, W., & He, S. (2015). *Vibrio Cholerae* Pathogen from The Freshwater-Cultured Whiteleg Shrimp *Penaeus vannamei* And Control with B dello *vibrio bacteriovorus*. *Journal of Invertebrate Pathology*, 130, 13-20.
- Costa AM, Buglione CC, Bezerra FL, Martins PCC and Barracco MA. 2009. Immune assessment of farm-reared *Penaeus vannamei* shrimp naturally infected by IMNV in NE Brazil. Aquaculture 291:141–146.
- Defoirdt T, Sorgeloos P, Bossier P. (2011). Alternatives to Antibiotics for The Control of Bacterial Disease In Aquaculture. Current Opinion in Microbiology 14:251–258.
- Garcia-Carreno FL, Cota K and Navarrete del Toro MA. 2008. Phenoloxidase activity of hemocyanin in whiteleg shrimp *Penaeus vannamei* : conversion, characterization of catalytic properties, and role in postmortem melanosis. Journal of Agricultural and Food Chemistry 56: 6454– 6459.
- Ghowina, CH (2018). The Effectiveness of Spirulina Platensis in Artificial Feed Against Hemocytes of Vaname Shrimp (Litopenaeus Vannamei) Infected by Vibrio Harveyi (Doctoral Dissertation, Brawijaya University).
- Fariedah, F. (2010). Effect of Vibrio Alginolyticus Immunostimulant Outer Membrane Protein (OMP) and Vibrio Harveyi Infection on Tiger Shrimp Mitochondrial DNA Penaeus Monodon Fab (Doctoral Dissertation, Universitas Brawijaya).
- Febriani, D., & Nuryati, S. (2013). Kappa-Carrageenan as an Immunostimulant for Controlling Infectious Myonecrsis (IMNV) in Vaname Shrimp, *Litopenaeus Vannamei*. *Journal of*

Indonesian Aquaculture , 12 (1), 70-78.

- (KKP) Ministry of Maritime Affairs and Fisheries. (2010). Aquaculture Production Target. Http://Www.Perikanan-Budidaya.Kkp.Go.Id. (June 23, 2011)
- (OIE) Office International of Epizootics. (2009). Manual of Diagnostics Test for Aquatic Animals: Infectious Myonecrosis. http://www.oie.int.eng/norms/fmanual/2.2.03_IMN.Pdf.(28 November 2010).
- Hernandez-Lopez, J., Gollas-Galvan, T., Vargas-Albores, F., 1996. Activation of the prophenoloxidase system of the brown shrimp Ž. *Penaeus californiensis* Holmes. Comp. Biochem. Physiol. 113C, 61–66.
- Jasmanindar Y. 2009. Used of *Gracilaria verrucosa* extract to increase white shrimp *Litopenaeus vannamei* defense system [Thesis]. Bogor (ID): Postgraduate School, Bogor Agricultural University.
- Kwang, LC, 1996. Immune Enhancer in the Control of Disease in Aquaculture. Encap Technology Pte. Ltd. Singapore.
- Lestari, NPT, Julyantoro, PGS, & Suryaningtyas, EW (2018). Challenge Test of Vibrio Harveyi Bacteria on Post Larvae of Vaname Shrimp (*Litopenaeus vannamei*). *Current Trends in Aquatic Science*, 1 (1), 114-121.
- Li F, Xiang J. 2013. Signaling pathways regulating innate immune responses in shrimp. Fish and Shellfish Immunology 34: 973–980.
- Loh, PC, Tapay, LM, Lu, Y., Nadala, EC, Jr., 1997. Viral pathogens of the penaeid shrimp. Adv. Virus Res. 48, 263–312.
- Muahiddah, N., & Sulystyaningsih, ND (2021). Analysis of Extraction Results of *Sargassum* Sp. From Teluk Ekas, the Trigger for Increasing Seaweed Production, East Lombok, West Nusa Tenggara. *Agroqua Journal: Agronomy and Aquaculture Information Media*, 19, 131-142.
- Omori, SA, Martin, GG, Hose, JE, 1989. Morphology of haemocyte lysis and clotting in the ridgeback prawn, Sicyonia ingentis. Cell Tissue Res. 255, 117–123.
- Pebrianto, CA (2009). Potential of Trichoderma Sp. As an Antibacterial and Immunostimulating Material in Vaname Shrimp, *Litopenaeus vannamei*.
- Permanti, YC, Julyantoro, PGS, & Pratiwi, MA (2018). The Effect of Adding Bacillus Sp. Against Vibriosis Infected Post Larvae of Shrimp vannamei (*Litopenaeus vannamei*). *Aquatic Sci*, 1 (1), 91-97.
- Rodriguez J, Moulac GL. 2000. State of the art of immunological tools and health control of penaeid shrimp. Aquaculture 191:109–119.
- Sakai, M. (1999). Current Research Status of Fish Immunostimulants. *Aquaculture*, 172 (1-2),63-92.
- Setyawan, A. (2019). Fucoidan From Tropical Brown Algae As An Immunostimulant In Vannamei
 Shrimp (*Litopenaeus Vannamei*): Hematological Studies, Immune Gene Expression,
 Resistance To WSSV, And Growth (Doctoral Dissertation, Gadjah Mada University).
- Siregar AF, Sabdono A, Pringgenies D. 2012. Antibacterial potential of seaweed extract against skin disease bacteria *Pseudomonas aeruginosa*, *Staphylococcus* epidermidis, and Micrococcusluteus. Journal of Marine Research 1:152–160.
- Sirirustananun N, Chen JC, Lin YC, Yeh ST, Liou CH, Chen LL, Sim SS, Chiew SL. 2011. Dietary administration of a *Gracilaria tenuistipitata* extract enhances the immune response and resistance against *Vibrio alginolyticus* and white spot syndrome virus in the white shrimp *Litopenaeus vannamei*.
- Soderhall, K., Cerenius, L., 1992. Crustacean Immunity. Annu. Rev. Fish Dis., 3–23.

- Surur, M. (2018). The Effect of Feeding Supplemented Sodium Alginate Extract on the Immune Response of Vannamei Shrimp (*Litopenaus vannamei*) (Doctoral Dissertation, Unisnu Jepara).
- Vargas-Albores, F., Hernandez-Lopez, J., Gollas-Galvan, T., Montano-Perez, K., Jimenez-Vega, F., Yepiz- ´ ~ Plascencia, G., 1998. Activation of shrimp cellular defense functions by microbial products. In: Flegel, T. Ž. ed., Advances in Shrimp Biotechnology. National Center for Genetic Engineering and Biotechnology, Bangkok, pp. 161–166.
- Wahjuningrum, D., Efianda, TR, Tarman, K., Yuhana, M., Effendi, I., & Saputra, F. (2020).
 Supplementation of *Nodulisporium* Sp. KT29 Induced by *Vibrio harveyi* As an Immunostimulant For Controlling Vibriosis in vannamei White Shrimp Under Marine Culture System. Journal of Indonesian Aquaculture, *19* (2), 95-105.
- Widanarni, Widagdo P, Wahjuningrum D. 2012. Oral Application of Probiotic, Prebiotic, And Synbiotic In Pacific White Shrimp *Litopenaeus Vannamei* Challenged with *Vibrio harveyi*. Journal of Indonesian Aquaculture 11: 54–63.
- Wijesekara I, Pangestuti R, Kim SK. 2011. Biological Activities and Potential Health Benefits of Sulfated Polysaccharides Derived from Marine Algae. Carbohydrate Polymers 84:14–21.
- Xu D, Liu W, Alvarez A, Huang T. 2014. Cellular immune responses against viral pathogens in shrimp. Developmental and Comparative Immunology 47: 287–297.
- Zahra, A., Sukenda, S., & Wahjuningrum, D. (2017). Extract of Seaweed *Gracilaria verrucosa* as Immunostimulant to Controlling White Spot Disease in Pacific White Shrimp *Litopenaeus vannamei*. Journal of Indonesian Aquaculture, 16(2), 174-183.